

Lead Optimization of Imidazopyrazines: A New Class of Antimalarial with Activity on *Plasmodium* Liver Stages

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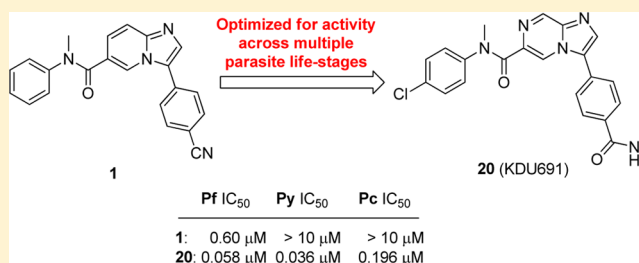
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S Supporting Information

ABSTRACT: Imidazopyridine **1** was identified from a phenotypic screen against *P. falciparum* (Pf) blood stages and subsequently optimized for activity on liver-stage schizonts of the rodent parasite *P. yoelii* (Py) as well as hypnozoites of the simian parasite *P. cynomolgi* (Pc). We applied these various assays to the cell-based lead optimization of the imidazopyrazines, exemplified by **3** (KAI407), and show that optimized compounds within the series with improved pharmacokinetic properties achieve causal prophylactic activity *in vivo* and may have the potential to target the dormant stages of *P. vivax* malaria.

KEYWORDS: Liver-stage antimalarial, imidazopyrazines, structure–activity relationship, lead optimization



Malaria continues to be a significant public health problem, threatening up to 40% of the global population.¹ Among the five species of malaria parasites that infect humans,² *P. vivax* malaria alone causes approximately 80–300 million clinical cases annually.^{3–5} Primaquine, the only licensed drug for the radical cure of *P. vivax*, is contraindicated in glucose-6-phosphate dehydrogenase (G6PD)-deficient populations,⁶ and the recommended long-term treatment (30 mg/kg for 14 days) precludes its widespread use. Thus, there remains an urgent need to develop non-8-aminoquinolines as safe and effective therapeutics for radical cure of *P. vivax*. From a drug discovery perspective, finding potential radical curative agents requires identifying compounds that also target the liver stages of the parasite.

We recently described two *in vitro* assays, which can identify compounds targeting developing schizonts and quiescent hypnozoites in hepatocytes.^{7–9} These assays utilize the rodent *P. yoelii* (Py) and the simian *P. cynomolgi* (Pc) parasites to target the liver schizonts and hypnozoites, respectively.¹⁰ We sought to develop a screening strategy involving both blood- and liver-stage assays to identify compounds with biological activity on multiple parasite life stages with the ultimate goal of developing radical curative agent for *P. vivax* malaria. A

subsequent candidate with optimal drug-like properties could then be evaluated in the *P. cynomolgi* infected monkey model as a test for radical cure activity.¹¹

From a previous phenotypic screening effort on Pf blood stages, we selected imidazopyridine **1** as a starting point despite its relatively weak activity (Figure 1).¹² Early hit-to-lead chemistry on the two distal aromatic rings resulted in the identification of **2**, a compound that showed a greater than 50-fold increase in potency. Although compound **2** was active on blood stages, it was still relatively inactive on Py schizonts and Pc and suffered from poor physicochemical properties, presumably due to the high lipophilicity (cLogP = 4.3). Modification of the bicyclic core by installation of a nitrogen atom at the 7 position reduced the cLogP by one log unit and resulted in **3** (KAI407).⁸ Although the potency on Pf blood stages was diminished, incorporating the imidazopyrazine core resulted in a compound with improved activity on Py and Pc liver stages.

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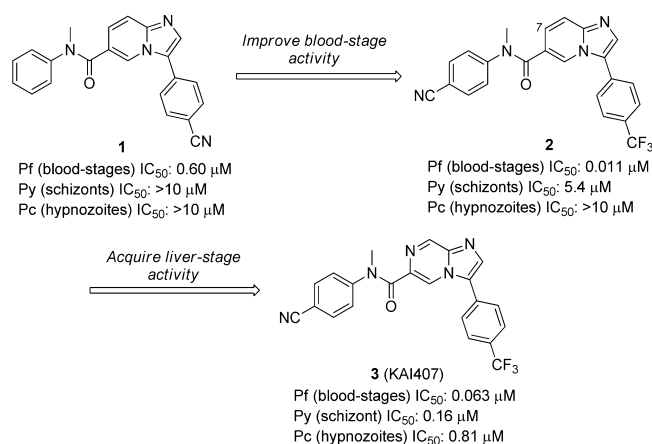


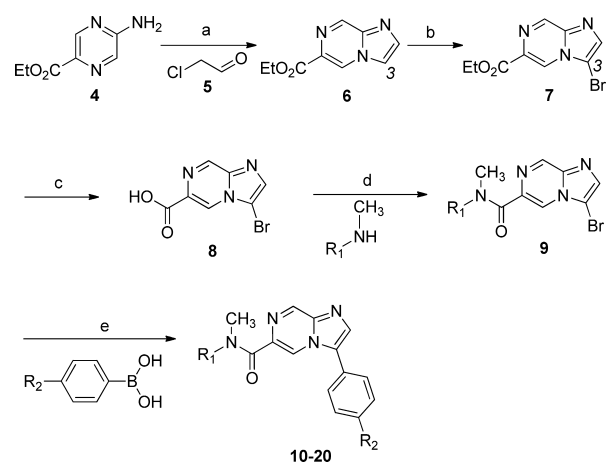
Figure 1. *In vitro* activity of compounds 1–3 on multiple parasite life stages.

The added activity of compound 3 on Py schizonts and Pc hypnozoites suggested that changes to the bicyclic core could impart the desired liver-stage activity to the molecule. We further hypothesized that activity in the Py assay might be predictive of potency on Pc hypnozoites for this chemical class and could therefore be used as a first-pass screen due to the low throughput of the Pc assay.

Following the discovery of compound 3, we initiated a lead optimization effort in an attempt to identify a compound with an optimal potency and PK profile. The overall strategy focused on making peripheral changes around the bicyclic imidazopyrazine core. This feature, along with the C3 phenyl and amide linker was found to be the main pharmacophore responsible for activity on Pc hypnozoites *in vitro*. Thus, analogues could be prepared from a primary building block consisting of the imidazopyrazine core (8) and chemically elaborated to 10–20 (Scheme 1).

Briefly, 2-chloroacetaldehyde 5 was added to a solution of aminopyrazine 4 in ethanol and heated to reflux to afford the

Scheme 1. Synthetic Route to the Imidazopyrazines^a



^aReagents and conditions: (a) 2-chloroacetaldehyde 5 (50 wt % solution in water), NaHCO₃, ethanol, reflux, 70%; (b) *N*-bromosuccinimide (NBS), CH₂Cl₂, room temperature (rt), 94%; (c) NaOH, THF/H₂O, 60 °C, 56%; (d) oxalyl dichloride, CH₂Cl₂, DMF (a drop), 30 min, followed by amine, rt; (e) boronic acid, Pd(PPh₃)₄ (5–10 mol %), aqueous KF solution (2 M), microwave, 110 °C, 30 min to 1 h.

imidazopyrazine core (6).¹³ A regioselective bromination at C3 with NBS provided 3-bromo-6-carboxylate 7 as the sole product isolated.¹⁴ Hydrolysis of 7 to the corresponding acid (8) followed by amide bond formation with an appropriate amine provided compound 9. Finally, Suzuki coupling with a series of boronic acids yielded the corresponding imidazopyrazine analogues.¹⁵

As we explored the SAR around the R₁ position of the imidazopyrazine core, we found that *para*-substituted *N*-phenyl or *N*-pyridyl were favored, while other heterocycles such as pyridazine (12) or benzothiazole (13) resulted in a loss of activity on both blood and liver stages (Table 1). In addition, the benzyl derivative (14), or replacement with a piperidine

Table 1. Blood- and Liver-Stage (Schizont) Activity of the Imidazopyrazines^a

cmpd	R ₁	R ₂	Pf IC ₅₀ (μM)	Py IC ₅₀ (μM)
3		-CF ₃	0.063	0.160
10		-CF ₃	0.275	0.200
11		-CF ₃	1.63	3.63
12		-CF ₃	> 10	> 10
13		-CF ₃	9.58	> 10
14		-CF ₃	> 10	> 10
15		-CF ₃	> 10	> 10
16			0.032	0.046
17			0.022	0.019
18		-CONH ₂	0.056	0.062
19		-CONHCH ₃	0.131	0.290
20		-CONHCH ₃	0.058	0.036

^aIC₅₀ values are the average of at least two independent experiments.

ring (**15**), resulted in a loss of potency suggesting a narrow range of tolerated substitutions at the R₁-position. The C3 phenyl was an essential part of the pharmacophore, and para-substitutions were preferred. Optimization of the R₂ groups revealed the carboxamide as the optimal substituent, providing a good balance between potency and physicochemical properties (**18–20**). Other R₂ substituents such as dimethylamine, methylsulfones, and carboxylic acids were inactive (data not shown). Interestingly adding an additional heterocycle such as the pyrazole (**16**) or aminothiadiazole (**17**) resulted in some of the most potent compounds. However, the poor solubility for these analogues led to low C_{max} and poor oral exposure in mice.

Only those compounds displaying blood- and liver-schizont activity (<300 nM) were selected for testing on Pc hypnozoites. This assay can also differentiate between the developing schizonts and dormant hypnozoites, which are not present in the Py assay. For this chemical series, we found that activity in the Py assay was predictive of activity on both Pc liver stages (Table 2).

Table 2. Activity of the Imidazopyrazines on *P. cynomolgi* Liver Stages^a

cmpd	<i>P. cynomolgi</i> IC ₅₀ (μM)	
	schizonts	hypnozoites
3	0.12	0.81
16	0.30	0.30
17	0.040	0.060
18	0.19	0.26
19	0.081	0.094
20	0.105	0.196

^aIC₅₀ values are the average of at least two independent experiments.

The specific physicochemical properties of the imidazopyrazines were largely dictated by the peripheral aromatic substituents (R₁ and R₂) (Table 3). In order to determine if

Table 3. Summary of Key Physicochemical Properties of the Imidazopyrazines

cmpd	solubility (μM, pH 6.8)	cLogP	mouse microsome stability	
			ER ^a (%)	T _{1/2} (min)
3	9	3.3	40	82
17	<5	1.9	na ^b	na
18	170	1.7	52	57
19	430	1.3	33	125
20	820	2.5	49	63

^aHepatic extraction ratio. ^bNot available.

Table 4. Pharmacokinetic Parameters Following 20 mg/kg oral and 5 mg/kg Intravenous Dosing in Mice^a

cmpd	oral PK ^b parameters					intravenous PK parameters		
	C _{max} (μM)	T _{max} (h)	AUC (μM·h)	T _{1/2} (h)	F (%)	V _{ss} (L/kg)	CL (mL/min/kg)	T _{1/2} (h)
17	0.02	1.7	0.12	3.39	0.24	0.99	16.2	2.12
18	8.3	0.5	11.14	1.35	51.6	0.90	39.7	0.62
19	5.2	0.5	12.4	1.63	51.9	1.02	34.0	0.42
20	20.5	0.5	31.2	3.63	60	0.96	19.3	0.83

^aC_{max}, maximum concentration of drug in plasma; T_{max}, time to maximum concentration of drug in plasma; AUC, area under the curve extrapolated to infinity; V_{ss}, volume of distribution at steady state; CL, clearance; T_{1/2}, half-life; F, oral bioavailability; ^bFormulation used for oral and intravenous dosing is PEG300/DSW (3:1, V/V).

the improved solubility would translate to a better *in vivo* PK profile, compounds **17–20** were evaluated in mice by oral (p.o.) and intravenous (i.v.) routes at 20 and 5 mg/kg, respectively (Table 4). Of the four compounds evaluated, **18** and **19** were more soluble and displayed favorable PK properties with moderate clearance, low volume of distribution, and moderate bioavailability (~50%). Increasing the clogP slightly by alkylating the primary carboxamide (**20**) further boosted C_{max} and exposure levels while lowering clearance and increasing the bioavailability (60%).

If Py liver schizonts are being inhibited *in vitro*, then the assay should predict that active compounds with an acceptable pharmacokinetic profile display causal prophylactic activity *in vivo* by preventing the parasite from establishing itself in the liver. In order to evaluate the *in vivo* efficacy of this class of compounds, analogues **3** and **18–20** were tested in a causal prophylaxis *P. berghei* mouse model (Figure 2). When

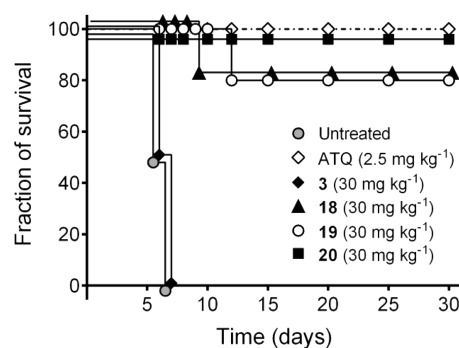


Figure 2. Single dose mouse causal prophylactic activity of the imidazopyrazines.

administered orally at a single dose of 30 mg/kg at the time of infection, compounds **18** and **19** were able to completely protect 80% of mice as compared to untreated control or compound **3**-treated. More attractively, compound **20** (KDU691)¹⁶ displayed comparable activity to atovaquone (ATQ) and was able to completely prevent a developing liver-stage infection; an indication that developing schizonts were being inhibited and precluding blood-stage patency.

In conclusion, we described the SAR around the imidazopyrazine class of antimalarials by exploring substitutions at the R₁- and R₂-substitutions of the bicyclic core. Optimization of pharmacokinetic properties by improving the solubility led to the identification of a series of compounds for *in vivo* efficacy evaluation. Compound **20** (KDU691) combined the optimal potency and PK properties to provide 100% protection from a developing *P. berghei* infection in a causal prophylaxis model at a single oral dose.

The target of the imidazopyrazines was recently identified and reported as Plasmodium PI4K, which we believe is the source of the broad antimalarial activity across the various parasite life stages.¹⁶ At the time of the lead optimization, this target was suspected, but recombinant protein was unavailable, and hence, the optimization was primarily driven by cellular activity in three different parasite assays. We have shown that the imadazopyrazines with dual activity on blood stages and liver schizonts also translated to potency on hypnozoites *in vitro*. Although the activity of **20** on liver-stage schizonts was confirmed in a casual prophylaxis mouse model, the test for *in vivo* antihypnozoite activity will require the *P. cynomolgi* infected monkey model for validation.

■ ASSOCIATED CONTENT

Supporting Information

Full experimental details for compounds synthesized and description of assays. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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Notes

The authors declare no competing financial interest.

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■ REFERENCES

- (1) World Health Organization. World Malarial Report 2011. http://www.who.int/malaria/world_malaria_report_2011/en/index.html.
- (2) White, N. J. Plasmodium knowlesi: the fifth human malaria parasite. *Clin. Infect. Dis.* **2008**, *46*, 172–173.
- (3) Mendis, K.; Sina, B. J.; Marchesini, P.; Carter, R. The neglected burden of plasmodium vivax malaria. *Am. J. Trop. Med. Hyg.* **2001**, *64*, 97–106.
- (4) Hay, S. I.; Guerra, C. A.; Tatem, A. J.; Noor, A. M.; Snow, R. W. The global distribution and population at risk of malaria: past, present, and future. *Lancet Infect. Dis.* **2004**, *4*, 327–336.
- (5) Mueller, I.; Galinski, M. R.; Baird, J. K.; Carlton, J. M.; Kochar, D. K.; Alonso, P. L.; del Portillo, H. A. Key gaps in the knowledge of *Plasmodium vivax*, a neglected human malaria parasite. *Lancet Infect. Dis.* **2009**, *9*, 555–566.
- (6) Alving, A. S.; Carson, P. E.; Flanagan, C. L.; Ickes, C. E. Enzymatic deficiency in primaquine-sensitive erythrocytes. *Science* **1956**, *124*, 484–485.
- (7) Meister, S.; Plouffe, D. M.; Kuhen, K. L.; Bonamy, G. M.; Wu, T.; Barnes, S. W.; Bopp, S. E.; Borboa, R.; Bright, A. T.; Che, J.; Cohen, S.; Dharia, N. V.; Gagaring, K.; Gettayacamin, M.; Gordon, P.; Groessl, T.; Kato, N.; Lee, M. C.; McNamara, C. W.; Fidock, D. A.; Nagle, A.; Nam, T. G.; Richmond, W.; Roland, J.; Rottmann, M.; Zhou, B.; Froissard, P.; Glynne, R. J.; Mazier, D.; Sattabongkot, J.; Schultz, P. G.; Tuntland, T.; Walker, J. R.; Zhou, Y.; Chatterjee, A.; Diagana, T. T.; Winzeler, E. A. Imaging of plasmodium liver stages to drive next-generation antimalarial drug discovery. *Science* **2011**, *334*, 1372–1377.

- (8) Zeeman, A. M.; van Amsterdam, S. M.; McNamara, C. W.; Voorberg-van der Wel, A.; Klooster, E. J.; van den Berg, A.; Remarque, E. J.; Plouffe, D. M.; van Gemert, G. J.; Luty, A.; Sauerwein, R.; Gagaring, K.; Borboa, R.; Chen, Z.; Kuhen, K.; Glynne, R. J.; Chatterjee, A. K.; Nagle, A.; Roland, J.; Winzeler, E. A.; Leroy, D.; Campo, B.; Diagana, T. T.; Yeung, B. K. S.; Thomas, A. W.; Kocken, C. H. KAI407, a potent non 8-aminoquinoline compound that kills *Plasmodium cynomolgi* early dormant liver stage parasites *in vitro*. *Antimicrob. Agents Chemother.* **2013**, *58*, 1586–1595.

- (9) Dembele, L.; Franetich, J. F.; Lorthiois, A.; Gego, A.; Zeeman, A. M.; Kocken, C. H.; Le Grand, R.; Dereuddre-Bosquet, N.; van Gemert, G. J.; Sauerwein, R.; Vaillant, J. C.; Hannoun, L.; Fuchter, M. J.; Diagana, T. T.; Malmquist, N. A.; Scherf, A.; Snounou, G.; Mazier, D. Persistence and activation of malaria hypnozoites in long-term primary hepatocyte cultures. *Nat. Med.* **2014**, *20*, 307–312.

- (10) *P. cynomolgi* has historically been used as a surrogate for *P. vivax* in a disease monkey model as both simian and human species form dormant liver-stages *in vivo*.

- (11) Schmidt, L. H. Appraisals of compounds of diverse chemical classes for capacities to cure infections with sporozoites of *Plasmodium cynomolgi*. *Am. J. Trop. Med. Hyg.* **1983**, *32*, 231–257.

- (12) Plouffe, D.; Brinker, A.; McNamara, C.; Henson, K.; Kato, N.; Kuhen, K.; Nagle, A.; Adrián, F.; Matzen, J. T.; Anderson, P.; Nam, T. G.; Gray, N. S.; Chatterjee, A.; Janes, J.; Yan, S. F.; Trager, R.; Caldwell, J. S.; Schultz, P. G.; Zhou, Y.; Winzeler, E. A. *In silico* activity profiling reveals the mechanism of action of antimalarials discovered in a high-throughput screen. *Proc. Natl. Acad. Sci. U.S.A.* **2008**, *105*, 9059–9064.

- (13) Chezal, J. M.; Moreau, E.; Delmas, G.; Gueiffier, A.; Blache, Y.; Grassy, G.; Lartigue, C.; Chavignon, O.; Teulade, J. C. Heterocyclization of functionalized vinylic derivatives of imidazo[1,2-a]pyridines. *J. Org. Chem.* **2001**, *66*, 6576–6584.

- (14) Chuaqui, C.; Cossrow, J.; Dowling, J.; Guan, B.; Hoemann, M.; Ishchenko, A.; Jones, J. H.; Kabigting, L.; Kumaravel, G.; Peng, H.; Powell, N.; Raimundo, B.; Tanaka, H.; Van Vloten, K.; Vessels, J.; Xin, Z. Preparation of heteroaryl compounds useful as Raf kinase inhibitors. WO2010/078408 A1.

- (15) Note that a weakly basic solution of aqueous potassium fluoride (KF) was required to avoid cleavage of the amide bond under the microwave conditions of the Suzuki reactions. Alternatively the order of the Suzuki and amide bond formation can be reversed.

- (16) McNamara, C. W.; Lee, M. C. S.; Lim, C. S.; Lim, S. H.; Roland, J.; Nagle, A.; Simon, O.; Yeung, B. K. S.; Chatterjee, A. K.; McCormack, S. L.; Manary, M. J.; Zeeman, A. M.; Dechering, K. J.; Kumar, T. R. S.; Henrich, P. P.; Gagaring, K.; Ibanez, M.; Kato, N.; Kuhen, K. L.; Rischli, C.; Rottmann, M.; Plouffe, D. M.; Bursulaya, B.; Meister, S.; Rameh, L.; Trappe, J.; Haasen, D.; Timmerman, M.; Sauerwein, R. W.; Suwanarusk, R.; Russel, B.; Renia, L.; Nosten, F.; Tully, D. C.; Kocken, C. H. M.; Glynne, R. J.; Bodenreider, C.; Fidock, D. A.; Diagana, T. T.; Winzeler, E. A. Targeting Plasmodium PI(4)K to eliminate malaria. *Nature* **2013**, *504*, 248–253.